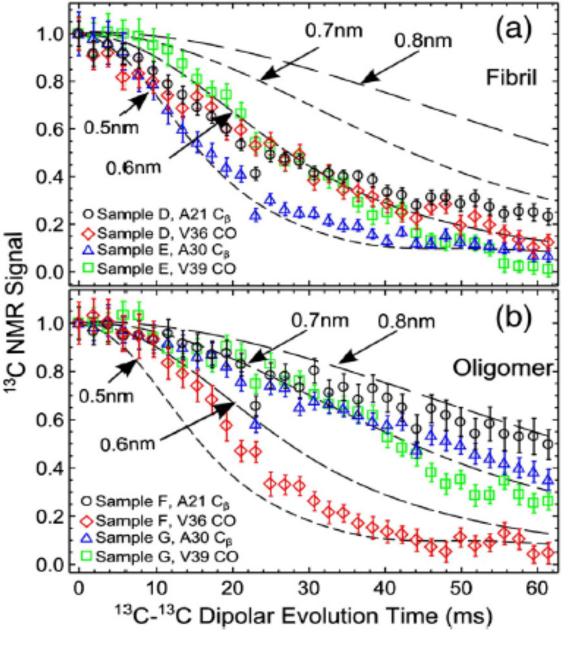
Solid State NMR Investigation of 150 kDa oligomers of the 42-residue Alzheimer's β-amyloid peptide

Donald Bush, Jr.¹, Anant K. Paravastu^{2,3} ¹ Lake Region High School, FSU RET participant, 1995 Thunder Rd, Eagle Lake, FL 33839 Solid State NMR method: Magic Angle spinning The sample (blue) is rotating with high B_0 frequency inside the main magnetic field (B_0) . The axis of rotation is tilted by the magic angle θ_m with respect to the direction of B_0 . In any condensed phase, a nuclear spin experiences a great number of interactions. The main three interactions often lead to very broad and featureless lines. By spinning the sample at the magic angle θ_m (ca. 54.74°, where $\cos^2\theta_m = 1/3$) with respect to the direction of the magnetic field, the normally broad lines become narrower, increasing the resolution for better MagicAngleSpinning<u>CC BY-SA 3.0</u> identification and analysis of the spectrum. Results 13C NMR chemical shifts are consistent with ordered β-strand structure 50 40 30 20 10 Two dimensional ¹³C-¹³C fpRFDR solid state NMR spectra of Sample A (IL4: Uniformly labeled ¹³C at I32, M35, G37, = 20· and V40). The off-diagonal crosspeaks represent single-bond correlations which allow residue-specific assignments within the ¹³C-labeled amino acids, as illustrated by colored lines. Positions of the offdiagonal peaks (chemical shifts) and linewidths indicate that the labeled residues are characterized by welldefined molecular structures, and that Ca 40 30 20 10 180 170 ¹³C NMR Frequency (ppm) and C β chemical shifts are consistent with β -strand molecular configurations. Results



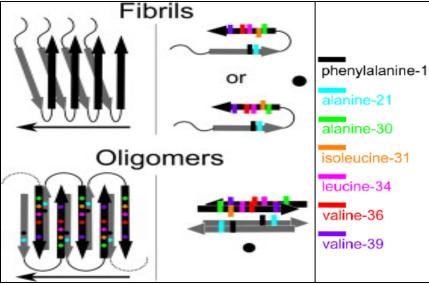
²National High Magnetic Field Laboratory, 1800 E. Paul Dirac Drive, Tallahassee, FL 32310 ³ Department of Chemical and Biomedical Engineering, FAMU-FSU College of Engineering, 2525 Pottsdamer Street, Tallahassee, FL 32310-6046 **Alzheimer's Disease and the β-amyloid peptide Paravastu lab previous A**β contributions Measurements of intermolecular ¹³C - ¹³C magnetic dipole-dipole couplings in C^{β} and Fibril CO for A β (1-42) fibril (a) and oligomer (b) 0.6 - 0.5nm samples by the PITHIRDS-CT solid state NMR technique. The lines represent predicted decays based on numerical simulations for linear strands of seven ¹³C nuclei equally spaced by the specified distances. Consistent decays for fibril samples labeled at multiple sites indicate an Oligomer in-register parallel β -sheet structure. Decreased decays for oligomer samples labeled at the A21 C^{β}, A30 C^{β}, and V39 CO O Sample F, A21 indicate that A β (1-42) oligomers are not Sample F, V36 CO Sample G, A30 C characterized by in-register parallel β -sheets.

Amyloid- β (A β) is the main component of amyloid plaques (extracellular deposits found in the brains of patients with Alzheimer's disease). Understanding amyloid- β $(A\beta)$ structures and underlying assembly pathways will advance the fundamental understanding of Alzheimer's disease (AD) at the molecular level. Recent research suggests that soluble oligomeric forms of Aβ may be causative agents in the development of Alzheimer's disease. It is generally believed that Aβ oligomers are the most toxic. A number of genetic, cell biology, biochemical and animal studies support the concept that A^β plays a central role in the development of Alzheimer's disease pathology, however there is no widely accepted structural model for Aβ oligomers. A more detailed understanding of Aβ oligomer assembly is likely to be an important step towards a molecular-level understanding of AD neurotoxicity. Recently presented a procedure for production of solid state NMR compatible 150 kDa oligometric samples of A β (1-42) (the 42-residue variant), and shown that oligometrs and amyloid fibrils differ in intermolecular arrangement of β -strands. O 0.8

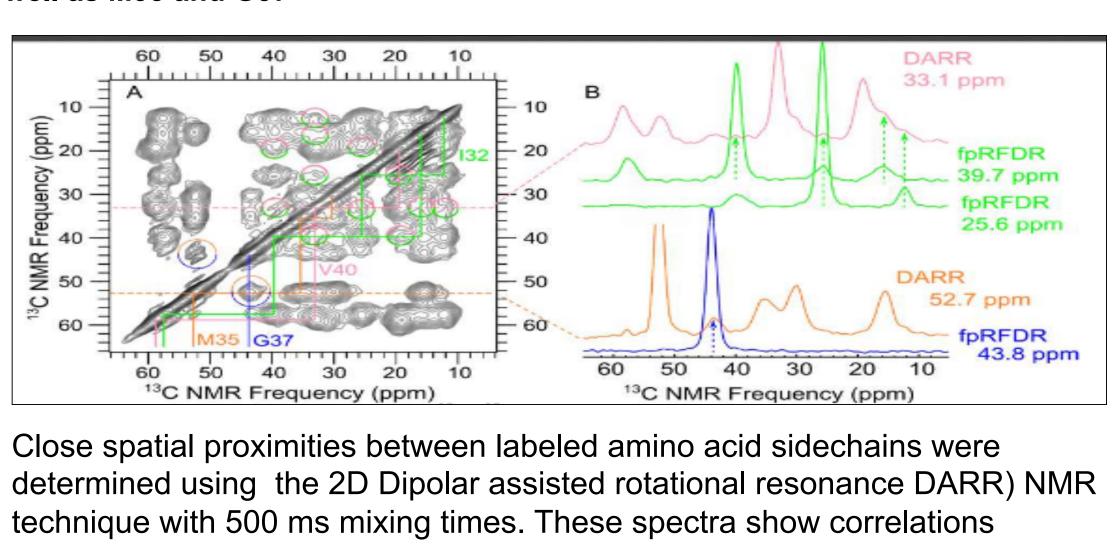


Hypothesis: A β 42 oligomers are arranged in antiparallel β -strands

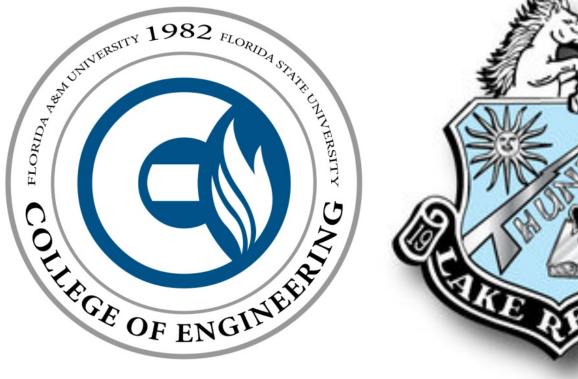
The most prominent structural differences between A $\beta(1-42)$ oligomers and fibrils were observed through measurements of intermolecular ¹³C–¹³C dipolar couplings observed in PITHIRDS-CT experiments. PITHIRDS-CT data indicate that, unlike fibrils, oligomers are not characterized by in-register parallel β -sheets.





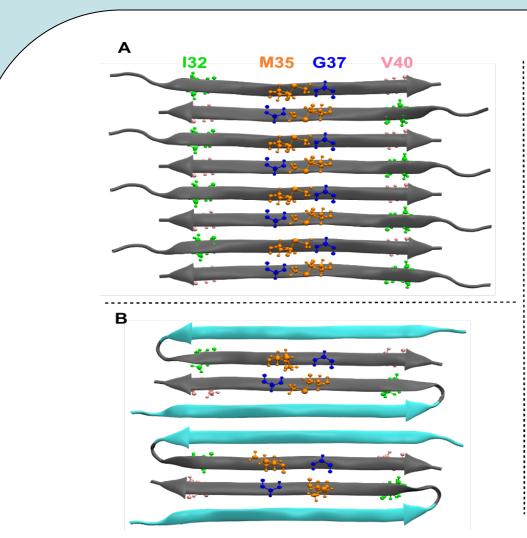


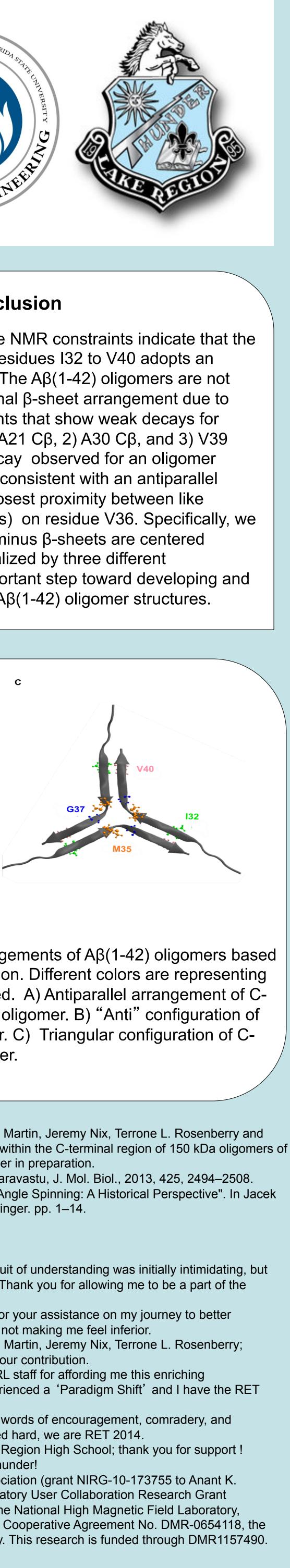
between atoms separated by distances up to ~0.6 nm. Analysis of the 2D DARR spectrum from Sample A (Figure A and 2B) indicates that the I32 sidechains are near the V40 sidechains and that the M35 sidechains are near the G37 sidechains.



Conclusion

For A β (1-42) oligomers, solid-state NMR constraints indicate that the approximate region spanned by residues I32 to V40 adopts an antiparallel β -sheet arrangement. The A β (1-42) oligomers are not thought to have a parallel C-terminal β -sheet arrangement due to previous PITHIRDS-CT experiments that show weak decays for three samples ¹³C –labeled on 1) A21 C β , 2) A30 C β , and 3) V39 CO. The strong PITHIRDS-CT decay observed for an oligomer sample ¹³C-labeled on V36 CO is consistent with an antiparallel C-terminal β-sheet "centered" (closest proximity between like residues on neighboring molecules) on residue V36. Specifically, we have shown the antiparallel C-terminus β -sheets are centered around V36. This finding is rationalized by three different molecular models which is an important step toward developing and testing hypothesis about 150kDa A β (1-42) oligomer structures.





Possible models of C-terminus arrangements of A β (1-42) oligomers based on the solid state NMR characterization. Different colors are representing different amino acid sites as illustrated. A) Antiparallel arrangement of Cterminus β -sheet within the A β (1-42) oligomer. B) "Anti" configuration of β -hairpins within in A β (1-42) oligomer. C) Triangular configuration of Cterminus β -sheet for A β (1-42) oligomer.

References

Danting Huang, Maxwell I. Zimmerman, Patricia K. Martin, Jeremy Nix, Terrone L. Rosenberry and Anant K. Paravastu "Antiparallel β-sheet structure within the C-terminal region of 150 kDa oligomers of the 42-residue Alzheimer's β -amyloid peptide" Paper in preparation. W. M. Tay, D. Huang, T. L. Rosenberry and A. K. Paravastu, J. Mol. Biol., 2013, 425, 2494–2508. Jacek W. Hennel, Jacek Klinowski (2005). "Magic Angle Spinning: A Historical Perspective". In Jacek Klinowski. New techniques in solid-state NMR. Springer. pp. 1–14.

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